

MICROCOPY RESOLUTION TEST CHART
URFAU Co. STANDARDS-1963-A



OFFICE OF NAVAL RESEARCH

Contract N00014-83-K-0470-P00003

Task No. NR 359-718

TECHNICAL REPORT # 44

In Situ Infrared Spectroelectrochemistry

Ву

Stanley Pons
J. Foley

Prepared for Publication in

Analytical Chemistry

University of Utah Department of Chemistry Salt Lake City. Utah 84112



July 30, 1986

Reproduction in whole or in part is permitted for any purpose of the United States Government.

This document has been approved for public release and sale; its distribution is unlimited.

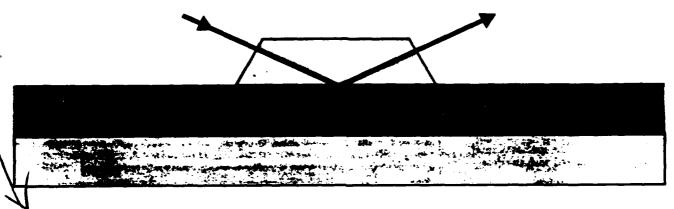
REPORT DOCUMENTATION PAGE		READ INSTRUCTIONS
		HEFORE COMPLETING FORM
HEPORT NUMBER 44	2. GOVT ACCESSION NO.	3. HECIPIEN I'S CATALOG NUMBER
In Situ Infrared Spectroelectroch	nemistry	5. TYPE OF REPORT & PERIOD COVERED
		Technical Report # 44
		6. PERFORMING ORG. REPORT NUMBER
Stanley Pons, J. Foley		6. CONTRACT OH GRANT NUMBER(s)
		N00014-83-K-0470-P0003
PERFORMING ORGANIZATION NAME AND ADDRESS		19. PROGRAM ELEMENT, PROJECT, YASK AREA & WORK UNIT NUMBERS
University of Utah Department of Chemistry Salt Lake City, UT 84112		Task No. NR 359-718
1. CONTROLLING OFFICE NAME AND ADDRESS	<u></u>	12. REPORT DATE
Office of Naval Research		July 30.1986
Chemistry Program - Chem Arlington, Virginia 22217	istry Code 472	13. NUMBER OF PAGES
4. MONITORING AGENCY NAME & ADDRESS(II dillored	nt from Controlling Office)	15. SECURITY CLASS. (of this report)
•		Unclassified
		15a. DECLASSIFICATION/DOWNGRADING
This document has been approved funlimited.		
7. DISTRIBUTION STATEMENT (of the abstract entered	in Dlock 20, If different free	n Report)
. SUPPLEMENTARY NOTES		
JR spectroelectrochemistry	-	intern C
An A-page article in Analytical C		ectroelectrochemistry.

Instrumentation

John K. Foley Stanley Pons

Department of Chemistry University of Utah Salt Lake City, Utah 84112

In Situ Infrared Spectroelectrochemistry



In situ infrared (IR) spectroelectrochemistry has become a viable experimental technique only within about the past five years, but it has already proved to be an important addition to the small number of in situ spectroelectrochemical methods currently in use (electron spin resonance spectrometry, ellipsometry, UV-visible spectrometry, resonance Raman, and surface-enhanced Raman spectrometry). The technique has found two broad areas of application.

 Adsorption on electrodes. In situ IR spectra can give information about the identity and molecular structure of adsorbed species and about the orientation and bonding of these species to the electrode surface.

• Electrode reactions. In situ IR spectra can conveniently be used to identify reactants, products, and intermediates of electrode reactions, both on the electrode surface and in solution. Species on the electrode surface can be distinguished from species in solution by varying the polarization state of the incident light.

In this article we will consider particularly the problems involved in obtaining IR spectra at the electrodeelectrolyte interface and how such problems are overcome by cell design and signal-processing techniques. We will also discuss how the spectra are affected by the nature of the electromagnetic electric fields near an electrode surface and the additional information about surface species that can be gained from this phenomenon. Finally, some examples of applications of IR spectroelectrochemistry will be given. We start with some general points about IR radiation and its absorption by molecules, which are pertinent to much of the following discussion.

If a molecule is to absorb IR radiation, two conditions have to be met. First, there must be an electromagnetic electric field where the molecule is situated, oscillating at the same frequency as a vibration of the molecule. The probability of absorption of radiation is proportional to the square of the strength of this field. Second, during the vibration of the molecule there must be a change in dipole moment with a component in the same direction as the electric field vector (the direction of oscillation of the field). The mechanism of absorption involves interaction between the oscillating dipole moment and the oscillating elec-

A beam of light incident on a plane surface (such as an electrode) can be considered to be made up of two com-

ponents, s-polarized light, which has its electric field vector oriented perpendicular to the plane of incidence (and therefore parallel to the surface) and p-polarized light, which has its electric field vector oriented parallel to the plane of incidence (and therefore with components both parallel to and perpendicular to the surface). Either s-polarized light or p-polarized light can be selected, if desired, by placing an appropriately oriented optical polarizer in the path of the beam.

Experimental problems

There are two major problems in measuring IR spectra of species at the electrode-electrolyte interface. The other in situ spectroelectrochemical techniques do not share these difficulties to the same extent and were consequently developed earlier. One problem, which occurs whenever IR radiation is passed through a solution. is absorption of most of the radiation by the bulk solvent. This is especially serious if the solvent is water, which absorbs strongly throughout most of the mid-IR region. The other problem is one of sensitivity. The number of molecules under study is usually very small (less than a surface monolaver in some experiments), while conventional IR sources are weak and IR detec-

0003-2700/85/0357-945A\$01.50/0 © 1985 American Chemical Society

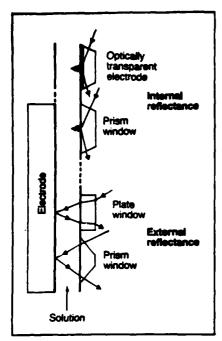


Figure 1. Optical geometries for internal- and external-reflection spectroelectrochemistry

tors are very noisy. This usually means that the total amount of radiation absorbed by the species of interest is small compared with the detector noise.

internal and external reflection

The most important factor in the design of an IR spectroelectrochemical cell is minimization of absorption of the incident IR radiation by the solvent. To achieve this, ideal electrochemical behavior has to be sacrificed. Two optical approaches have been used—internal reflection and external reflection.

The older method is attenuated total reflection (ATR) (1-3). In this method the light is passed through the cell at an angle such that it undergoes total internal reflection at the electrode-solution interface (Figure 1). The electric field of internally reflected light penetrates a short distance across the interface, its intensity falling off exponentially with distance into the solution. The depth of penetration of this "evanescent wave." which is a function of the wavelength of the radiation, is greatest at the critical angle and decreases to about one-tenth of the wavelength of the light at angles well above the critical angle. This means that the internally reflected radiation can be absorbed by species on the solution side of the interface, but only by species either adsorbed on the electrode or in solution within a micrometer or so of the electrode. Thus the bulk of solution does not contribute to the total absorbance.

This type of experiment puts quite

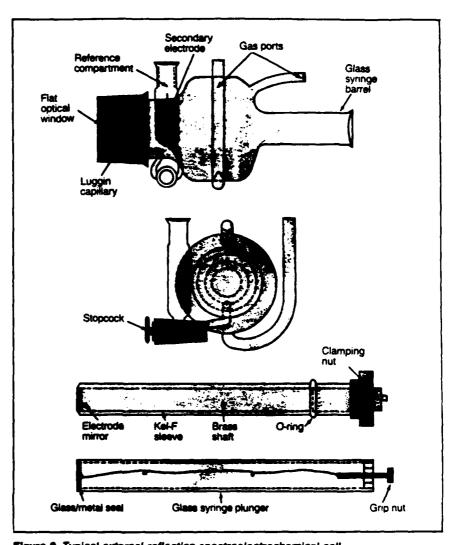


Figure 2. Typical external-reflection spectroelectrochemical cell. The top two drawings are side and end-on views of the cell. Either of the two mirror electrode assemblies at the bottom fits into the glass syringe barrel. The mirror is pushed forward until flat against the optical window.

stringent demands on the electrode material, which must both conduct electricity and transmit IR light. Because good conductors are also good IR absorbers, a compromise between these properties has to be reached. Early workers used germanium, which is an IR-transparent semiconductor. More recently, very thin films of metal (Au, Pt, or Fe) deposited onto IRtransparent substrates have been used. Near the critical angle, and if the solution is not too absorbing, the electric field in the solution adjacent to the electrode can be several times greater than the electric field in the incident beam. This enhancement is greater for the latter type of electrode than for the former. Both single-reflection and multiple-reflection arrangements have been designed.

The more common way of reducing solvent absorption is to use external reflection in a thin-layer cell (Figure 1). In external reflection one end of the cell is an IR-transparent window. The IR beam passes through this win-

dow and through the solution, is specularly reflected from a polished electrode surface, and travels back through the solution and the window to a detector. The electrode is positioned very close to the window so that only a very thin layer of solution is sandwiched between them. The working electrode is a disk sealed into the end of an insulating cylinder. which can be moved inside a syringe barrel oriented perpendicular to the window (Figure 2). For the IR spectroscopic experiments the electrode is pushed forward until it makes contact with the window, forming a thin-layer cell with solution thickness of 1-100 µm.

the expressions, as increase as as respected in the second respected as the aspect of the respect

The material for the IR-transparent window is chosen primarily for high transmission and insolubility in the solvent. Windows have been made from calcium fluoride, silicon, silica. Irtran 2, Irtran 4, and zinc selenide. Two shapes have been used—tlat and prismatic. Prismatic windows allow the beam to strike the electrode at

more glancing angles, which increases the sensitivity for adsorbed species. Also, if the incident beam is normal to the edge of the window, nearly equal amounts of s- and p-polarized light enter the cell, which is important if the difference in absorption of the two polarizations is to be used to distinguish surface species from solution species. Flat windows preferentially transmit p-polarized light.

The electrode material has to be able to specularly reflect IR radiation. Polished disks of Pt. Rh. Au, Ag, Pd, Fe, C, and p-type CdTe have all been used. However, even if the electrode does not reflect (as seems to be the case for carbon electrodes in contact with some aqueous solutions), it is still possible to obtain spectra of solution-free species by using a prismatic IR-transparent window with angles of incidence where internal reflection occurs at the window-solution interface (Figure 1).

IR spectroelectrochemical cells usually use the potentiostated three-electrode arrangement. However, both the ATR and external-reflection configurations suffer from unusually high resistances in the current path. In an ATR cell the high resistance is in the working electrode itself, which must have low conductivity if it is to transmit IR light. In an external-reflection cell the thin layer of solution between the working electrode and the window has the high resistance. The result is that when current flows the electrode potential differs by an amount iR from the value controlled by the potentiostat, and this difference increases across the electrode as the resistance in the current path does. Furthermore, the response of the cell to a change in potential is slow because the RC time constant for charging the double-layer capacitance is high.

Signal processing

Signal-processing techniques in in situ IR spectroelectrochemistry serve two purposes—to remove the spectrum of the bulk solvent, which would otherwise swamp out the spectrum of adsorbed or electrogenerated species, and to enhance the very low S/N ratios encountered in these experiments.

The first goal is nearly always achieved by potential modulation. The electrode potential is changed from a value E_1 where the reflectance is R_1 to a value E_2 where the reflectance, R_2 , is different. The quantity measured is a function of

$$\frac{\Delta R}{R}\left(\overline{v}\right) = \frac{R_2(\overline{v}) - R_1(\overline{v})}{R_1(\overline{v})}$$

and spectra are usually presented as plots of $\Delta R/R$ vs. wavenumber \tilde{v} . Thus the spectrum is a difference spectrum, which shows only changes in reflec-

tance caused by the change in potential. The spectra of the bulk solution, atmosphere, etc., generally do not change with potential and are canceled out in the subtraction. Note that spectra are also sometimes plotted as

% transmittance =
$$100 \left(\frac{R_2}{R_1} \right)$$

= $100 \left(\frac{\Delta R}{R} + 1 \right)$
or absorbance = $-\log_{10} \left(\frac{R_2}{R_1} \right)$
= $-\log_{10} \left(\frac{\Delta R}{R} + 1 \right)$

against wavenumber.

A change in potential can cause a change in the reflectance for any of several reasons, including changes in the number and bonding of species adsorbed on the electrode, migration of ions into or out of the optical path, electron transfer reactions that form or destroy films on the electrode surface, and electron transfer reactions that form or consume species in solution.

Several approaches to increasing the S/N ratio are in use. depending particularly on whether a dispersive or a Fourier transform IR (FT-IR) spectrometer is used. With a dispersive instrument the variation of reflectance with potential is measured at constant wavenumber, whereas with an FT-IR instrument the variation of reflectance with wavenumber is measured at constant potential. Use of a dispersive spectrometer therefore implies an ac potential program, whereas either dc or ac potential programs can be used with an FT-IR spectrometer.

When using a dispersive instrument the most common method is to modulate the electrode potential between E_1 and E_2 with a square wave (or sine wave) of about 10-Hz frequency, while changing the wavelength of the light very slowly. The signal from the IR detector then consists of a dc component, $R(\hat{v})$, a small ac component, $\Delta R(\bar{v})$, at the same frequency as the potential modulation, and noise over a wide range of frequencies. This signal is fed into a lock-in amplifier which picks out only that component of the signal at the same frequency as the alternating potential. This provides a direct measure of ΔR at each wavenumber and very effectively removes noise at other frequencies. For further discrimination against noise, the output of the lock-in amplifier can be stored in a computer, and a number of spectra can be signal averaged. The number required with modern commercially available instruments is quite small. S/N is increased with the square root of the number of signalaveraged spectra. The background

spectrum $R(\bar{\nu})$ is obtained by mechanically chopping the beam and using the lock-in amplifier to pick out the component of the detector signal at the chopping frequency. This spectrum is also stored in the computer, and $\Delta R/R(\bar{\nu})$ can then be calculated. When this method is used in an external-reflection experiment, it is known as electrochemically modulated IR spectroscopy (EMIRS) (4-6).

Another approach that has been used with a dispersive instrument is linear potential-sweep IR reflectance spectroscopy (LPSIRS) (7). In this case, at each wavenumber the potential is swept rapidly over the required range. The time-varying output of the detector, which is a measure of reflectance as a function of potential at constant wavenumber, is stored in a computer. Several of these sweeps can be signal averaged to increase the S/N. The experiment is carried out at several different wavenumbers, and a series of spectra of reflectance against wavenumber at constant potential is reconstructed from the data. One potential is taken as the reference potential. E_1 , and the spectrum at this potential, $R_1(\bar{v})$, is divided into each of the other difference spectra to give $R_2(\bar{v})/R_1(\bar{v})$, which can be converted to $\Delta R/R(\bar{v})$.

FT-IR spectrometers are being used in increasing numbers for IR spectroelectrochemical experiments because of their very high throughput and because of their ability to acquire large numbers of spectra rapidly, which follows from the simultaneous sampling of the entire range of wavelengths. In an FT-IR spectroelectrochemical experiment the electrode potential is modulated between E_1 and E_2 . At each potential a large enough number of interferograms are added to give the desired S/N, and the two resulting single-beam spectra are ratioed to give the difference spectrum $R_2(v)/R_1(v)$. S/N is proportional to the square root of the number of interferograms collected. The potential-time program may be either a simple potential step from E_1 to E_2 or a square wave of frequency low enough that an integer number of interferograms can be collected during the period that the potential is held at each value. The square wave program reduces the effects of long-term drift in the spectrometer output but is not highly suitable for very complicated or chemically irreversible processes. When used with an external-reflection cell, this method is known as subtractively normalized interfacial FT-IR spectroscopy (SNIFTIRS) (4, 5, 8, 9).

A variation of the SNIFTIRS method is polarization-modulation or FT-IRRAS (FT-IR reflection-absorption spectroscopy) (10). A photoelastic modulator is used to modulate the

DOPY INSPECTED

state of the light between s- and p-polarization at a frequency of about 70 kHz, depending on the photoelastic modulator, which is much higher than the modulation frequencies of the interferogram. A lock-in amplifier is used to pick out the component of the detector response at the same frequency as the polarization modulation, and the output of the lock-in amplifier is Fourier transformed to give a spectrum that is the difference between the spectrum from the p-polarized light and the spectrum from the s-polarized light. For a distance of about one-quarter wavelength of the light from the electrode surface, s-polarized light is absorbed less than ppolarized light. Therefore the polarization modulation difference spectrum is the spectrum of a region about 1 um thick adjacent to the electrode surface, which is about the thickness of the thin layer of solution between the electrode and window. To remove the spectrum of the solvent it is necessary to carry out the experiment at two potentials and ratio the spectra. as in a SNIFTIRS experiment. The main advantage of this technique is probably the increased sensitivity from the additional phase-sensitive detection stage.

Reflection spectroscopy of surface species

In an ATR experiment the electric fields of both s-polarized and p-polarized light penetrate the interface. At the critical angle the evanescent wave from p-polarized light has only a strong electric field perpendicular to the surface, whereas the evanescent wave from s-polarized light has only a weak electric field parallel to the surface. It has been pointed out (1) that differences in absorption of the two polarization states by an adsorbed molecule could be used to determine its orientation, but no experimental demonstrations using this technique have appeared. At other angles of incidence p-polarized light gives rise also to a weak electric field parallel to the surface. In general, therefore, p-polarized light interacts more strongly with absorbing species than does s-polarized light, and ATR experiments use either unpolarized or p-polarized radiation.

The situation is more interesting in an external-reflection experiment. Upon specular reflection at a metal surface, s-polarized light undergoes a phase shift of close to 180° at all angles of incidence, and so the incident and reflected beams interfere destructively. Therefore, the electric field of s-polarized light has zero intensity close to the surface, and species near the surface cannot absorb s-polarized light. On the other hand, p-polarized

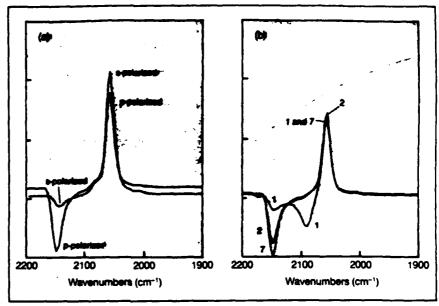


Figure 3. Difference SNIFTIRS spectra of the SCN⁻ system in acetonitrile at a silver electrode between -0.7 and +0.4 V vs. Ag/Ag⁺ reference using (a) p- and s-polarized radiation, (b) p-polarized radiation at 1 min, 2 min, and 7 min after potential pulse Vertical scales are 2 × 10⁻¹ \(\Delta R/R\) per division

light undergoes smaller phase shifts upon reflection, and there is a finite component of the electric field at the surface, but only in a direction normal to the surface. The strength of this electric field increases with angle of incidence, being zero at normal incidence and reaching a maximum close to glancing incidence. High angles of incidence are therefore advantageous if the spectra of adsorbed species are to be observed.

Two important and useful consequences follow from the presence at the surface of electromagnetic electric fields only normal to the surface and only arising from p-polarized radiation. First, species on the electrode surface can be distinguished from species in solution because species in solution can absorb both s-polarized and p-polarized light, whereas species on. or close to, the surface can absorb only p-polarized light. Second, IR radiation can interact only with those modes of vibration of an adsorbed molecule that have a component perpendicular to the surface of the change in dipole moment with normal coordinate. (This is the surface selection rule [11].)

Therefore, information about the orientation of an adsorbed molecule can be obtained by noting which of its modes of vibration are IR active and which are forbidden by the surface selection rule. To take a simple example, the C = O stretching vibration of an adsorbed CO molecule will give an external reflection IR spectrum with p-polarized light if the C = O bond is perpendicular to the surface, but not if the C = O bond is parallel to the surface. At no orientation will the molecule absorb s-polarized radiation. Re-

cent results. however, indicate that orientation data for very polarizable molecule: deduced from the surface selection rule should be treated with caution. There is a very high electrostatic electric field (106-108 V cm-1) at the electrode-electrolyte interface. normal to the electrode surface, which can induce a dipole moment normal to the surface in a sufficiently polarizable adsorbed molecule. Even though the permanent dipole of such a molecule may be oscillating parallel to the surface during a vibration (or even if there is no change in permanent dipole moment at all), the induced dipole moment can oscillate normal to the surface at the vibrational frequency, providing a mechanism for absorption of IR radiation (12). Bands observed from flat-adsorbed TCNE radical anion and acrylonitrile have been attributed to this effect.

Applications to electrode reactions

FT-IR spectrometers have been used more often than dispersive spectrometers to obtain spectra of the reactants and products of electrode reactions. There are some disadvantages to using dispersive instruments for these applications because of the acpotential programs that must be used. The short time scale of the ac experiment allows relatively little reaction to take place during each cycle, so changes in reflectance are small. If a chemically irreversible reaction is taking place in a thin-layer cell then all of the reactant in the thin layer may be consumed within the first few cycles of the potential, and no change in reflectance will be seen in the remainder of the spectrum. If the reaction mechanism is complicated, a complicated time-averaged mixture of species close to the electrode may be obtained, making the spectra difficult to interpret.

External-reflection methods seem to be superior to internal-reflection methods in the study of electrode reactions. External reflection uses a thin-layer cell, where bulk electrolyses of solution species can be quickly and conveniently carried out and the IR spectra of reactants and products obtained in a nearly ideal double-beam experiment. Spectra of intermediates can also be obtained if they are stable on the time scale of an FT-IR spectrum. Species on the electrode surface can be distinguished from species in solution by changing the polarization state of the radiation or noting potential-dependent band positions.

We use as an example anodic processes that take place at a silver electrode in acetonitrile solutions of thiocyanate ion, SCN- (13). Cyclic voltammetry shows two oxidation waves, the first characteristic of a reversible reaction with reactant and product in solution, the second characteristic of a surface process. SNIFTIRS spectra were taken with a single potential step from E_1 to E_2 . A step from a potential where no reaction occurs to a potential in the first wave gives a product absorbing at 2092 cm⁻¹, shown to be a solution-soluble species by its absorption of both s-polarized and p-polarized light, and thought to be the complex Ag(SCN)=2. Figure 3a shows the spectrum obtained in a potential step experiment from a potential where no reaction occurs to a potential just past the second voltammetric wave. The sign convention for $\Delta R/R$ has bands due to absorption by reactants at the base potential pointing up $(\Delta R/R > 0)$

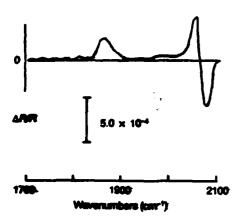


Figure 4. Difference EMIRS spectrum for methanol adsorbed on a platinum electrode from 1 M H₂SO₄ solution Modulation +0.06 to +0.45 V (vs. NHE) at 8.4 Hz

and bands due to products pointing down $(\Delta R/R < 0)$. It can be seen that a solution-soluble species absorbing at 2056 cm⁻¹, which is presumably SCN-, is converted to a species that absorbs only p-polarized light at 2141 cm⁻¹. This surface product is thought to be a film of AgSCN. Figure 3b shows spectra taken with the same potentials as in 3a (this time using only p-polarized light) with the spectra at the more positive potential taken at various time intervals after the potential step. This shows that SCN-(at 2056 cm-1) is initially converted to Ag(SCN)₂⁻ (at 2092 cm⁻¹), which later forms AgSCN on the surface (at 2141 cm⁻¹). The reactions can therefore be written

First wave:

 $Ag(s) + 2SCN^- = Ag(SCN)_2^- + e^-$ Second wave:

$$Ag(s) + 2SCN^- = Ag(SCN)_2^- + e^-$$

 $Ag(SCN)_2^- + Ag(s) = 2AgSCN(s) + e^-$

The most successful use of the ATR method has been in the study of polymer films on electrodes, where very high concentrations of absorbing species exist close to the electrode surface. A system that has been studied by FT-ATR is poly(3-methylthiophene) on a thin film of gold (3). Trnsitions between the semiconducting neutral species and the highly conducting oxidized state were clearly indicated in the IR spectra.

Thin layers of iron on germanium were used by the same workers to study the oxidation products of iron in alkaline media.

Applications to surface studies

All IR spectroelectrochemical studies of adsorbed species have been carried out by external-reflection methods. EMIRS has so far proved to be the most sensitive of these methods, but because of the ac potential program used, EMIRS spectra are also the most difficult to interpret.

A particularly interesting EMIRS study was carried out on the oxidation of methanol on platinum in aqueous acid solutions (14). The spectrum obtained by modulating between two potentials where methanol is adsorbed is shown in Figure 4. Bands pointing up $(\Delta R/R > 0)$ correspond to species present at the more negative potential. There is a weak band at 1850 cm⁻¹, attributed to CO adsorbed in a site where it is bridge bonded to three platinum atoms, and a strong bipolar band at about 2050 cm⁻¹, attributed to linearly adsorbed CO (i.e., bonded to one Pt atom). The bipolar nature of this band is caused by a shift in band position to more positive wavenumbers as the electrode potential is made

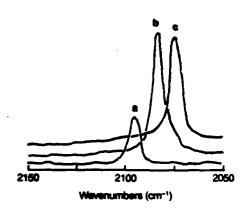


Figure 5. FT-IRRAS spectra of CO adsorbed on a platinum electrode from 1 M HCIO₄ solution

These are difference spectra taken between 0 8 V and (a) 0.675 V, (b) 0.4 V, and (c) 0.0 V (vs. NHE). The maximum absorbance is \sim 5 \times 10⁻²

more positive. This shift is most probably the result of strengthening of the C = 0 bond by withdrawal of electrons from the C = 0 antibonding π^* orbitals when the electrode is made more positive.

Because of its importance in catalysis and because it has a very intense band at a frequency well away from the regions of strong water absorption. linearly adsorbed CO has been studied a great deal. Figure 5 shows FT-IRRAS spectra of CO adsorbed on Pt in 1 M HClO₄ (15). For all of the spectra the reference potential $E_1 = 0.8 \text{ V}$. where no CO is adsorbed. The shift of band position to higher wavenumber with increasing positive potential can easily be seen, together with a decrease in intensity at higher potentials due to oxidation to CO2 of some of the adsorbed CO.

An example of the use of the surface selection rule to predict the orientation of an adsorbed molecule is provided by the case of p-difluorobenzene adsorption on Pt from aqueous acid solution (16). This molecule has 13 IRactive normal modes, of which 10 (the biu and biu modes) have dipole changes in the plane of the aromatic ring, and three (the big modes) have dipole changes perpendicular to the plane of the ring. If the molecule is adsorbed flat, only the bay modes should be seen, whereas if it is adsorbed edgewise only the blu and blu modes should be seen. At intermediate ("skew") orientations all modes should be seen. SNIFTIRS spectra with modulation between a potential where p-difluorobenzene is adsorbed and a potential where it is not adsorbed show, at concentrations less than 0.8 mM, two bands due to the ad-

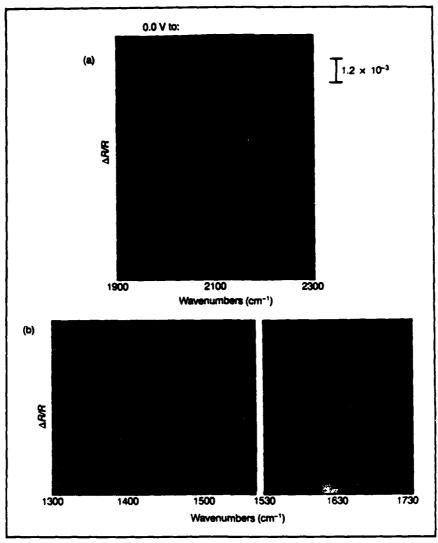


Figure 6. EMIRS difference spectra for adsorption of benzonitrile on a gold electrode from H_2SO_4 solution

Modulation at 8.5 Hz. (a) C \equiv N stretch region, voltage limits as indicated vs. SCE. (b) Ring mode region, voltage limits 0 to 1.1 V vs. SCE

sorbed species corresponding to two of the three b_{3u} modes. At concentrations above 0.8 mM, bands due to all of the modes appear. Therefore, at solution concentrations below 0.8 mM, p-difluorobenzene is adsorbed flat on the electrode, but a transition to skew orientation takes place at higher solution concentrations.

In addition, benzonitrile and acrylonitrile adsorbed on gold electrodes from aqueous acid solutions are examples of systems in which the surface selection rule is respectively successful and unsuccessful in predicting the orientations of adsorbed molecules (17).

Figure 6 shows EMIRS spectra taken in a solution of benzonitrile (0.02 M) and H₂SO₄ (1 M) at a good electrode. Modulation from a fixed base potential to more positive potentials gives a negative-going band at about 2250 cm⁻¹, which shifts to higher wavenumber with increasing positive potential. The sign of the band

and the potential dependence of its intensity indicate increasing adsorption of benzonitrile with increasing positive potential, until at very positive potentials formation of gold oxide and oxidation of benzonitrile take place. There can also be seen a very small positive-going band at 2229 cm⁻¹ arising from solution-free benzonitrile, which is depleted at the positive modulation limit because of its adsorption onto the electrode. In other regions of the spectrum there are two negativegoing bands at 1480 cm⁻¹ and 1595 cm⁻¹, corresponding to A₁ benzene ring modes, and a weak partive-going band at 1448 cm⁻¹, corre-nding to a B₁ ring mode. These obsections are all consident with benzonstrile adsorbed perpendicular to the surface and bonded to the surface through the N atom. Bonding through the N atom might be expected to involve donation of electrons from the slightly antibonding nitrogen lone pair orbital to

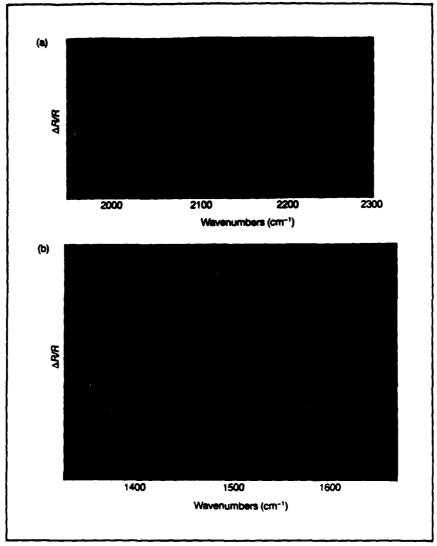


Figure 7. EMIRS difference spectra of acrylonitrile adsorbed on a gold electrode in (a) the C = N stretch and (b) the C = C stretch regions

Modulation from 0.05 to 1.0 V vs. SCE at 8.5 Hz

empty metal orbitals and back donation of electrons from filled metal orbitals to empty benzonitrile π^* orbitals. For both bonding mechanisms, a shift of the electrode potential to more positive values would strengthen the $C \equiv N$ bond and would shift the $C \equiv N$ stretch mode to higher wavenumber, as observed. The surface selection rule also predicts orientation of the $C \equiv N$ bond normal to the surface, because a strong $C \equiv N$ stretch band and bands from A_1 ring modes are observed.

200 X

An EMIRS spectrum from a solution of acrylonitrile (0.05 M) and $\rm H_2SO_4$ (1 M) at a gold electrode is shown in Figure 7. Modulation to a more positive potential gives positivegoing bands at about 2130 cm⁻¹ and 1520 cm⁻¹, with potential-dependent frequencies and intensities. These bands are assigned to C \approx N and C = C stretching vibrations, respectively, but occur about 100 cm⁻¹ lower in frequency than the corresponding

bands of solution-free acrylonitrile. This is evidence that the molecule is adsorbed flat, where it would probably be bonded to the electrode by donation of acrylonitrile n-bonding electrons to empty orbitals on the metal and by back donation of electrons from filled metal d-orbitals to empty acrylonitrile π^* orbitals. In this case. the C = N and C = C bonds would be weakened by adsorption, and the C = N and C = C stretch modes of the adsorbed molecule would be at lower wavenumbers than the corresponding modes of the solution-free molecule. Flat orientation of the adsorbed molecule is not predicted by a simple application of the surface selection rule, because C = N and C = C bands are observed. However, these bands are very weak, and the C = C band is stronger than the C = N band, whereas the opposite is the case for the free molecule. The most likely explanation for the appearance of these bands is the pres-

ence normal to the surface of a molecular dipole moment induced by the high electric field across the double laver. As the molecule vibrates, this dipole can oscillate normal to the surface and interact with the electric field of p-polarized IR radiation.

It is clear that IR spectroelectrochemistry can provide valuable insight into the vibrational structure of surface-bound and solution-soluble intermediates. Extending the technique to the far-IR to observe metal-adsorbate structure and use of the technique for surface dynamics studies are likely short-term extensions of the tech-

We would like to acknowledge the Office of Naval Research for support of part of this work.

References

- (1) Hansen, W. N. In "Advances in Electrochemistry and Electrochemical Engi-
- trochemistry and Electrochemical Engineering"; Muller, R. H., Ed.; Wiley: New York, N.Y.; 1973; Vol. 9, pp. 1-60.

 (2) Lange, P.: Glaw, V.: Neff, H.: Piltz, E.; Sass, J. K. Vacuum 1983, 33, 763.

 (3) Neugebauer, H.: Nauer, G.; Neckel, A.; Tourillon, G.; Garnier, F.; Lange, P. J. Phys. Chem. 1984, 88, 652.
- (4) Bewick, A.; Pons, S. In "Advances in
- (4) Bewick, A.; Pons, S. In "Advances in Infrared and Raman Spectroscopy";
 Hester, R.J.H.; Clark, R. E., Eds.; Wiley-Hayden: London, U.K., 1985; p. 1.
 (5) Pons, S.; Foley, J. K.; Russell, J.; Seversen, M. In "Modern Aspects of Electrochemistry"; Bockris, J. O'M.; Yeager, E., Eds.; Wiley: New York, N.Y., in press.

- (6) Bewick, A.; Kunimatsu, K.; Pons, B. S.; Russell, J. W. J. Electroanal. Chem. 1984, 160, 47.
- (7) Kunimatsu, K. J. Electroanal. Chem. 1983, 145, 219.
- (8) Pons, S.: Davidson, T.: Bewick, A. *J. Electroanal Chem.* 1984, 160, 63. (9) Habib, M. A.. Bockris, J. O'M. *J. Elec-*
- troanal. Chem. 1984, 180, 287. (10) Seki, H.; Kunimatau, K.; Golden, W. G., submitted for publication in
- Appl. Spectrosc. (11) Greenler, R. G. J. Chem. Phys. 1966. 44. 310.
- (12) Korzeniewski, C.; Shirts, R. B.; Pons, S.; J. Phys. Chem. 1985, 89, 2297
- (13) Foley, J. K.; Pons, S.; Smith, J. J., submitted for publication in Langmuir.
 (14) Beden, B.: Bewick, A.; Kunimatsu, K.;
- Lamy, C. J. Electroanal. Chem. 1983,
- 121, 343. (15) Kunimatsu, K.; Golden, W. G.; Seki, H.; Philpott, M. R. Langmuir 1985, 1,
- (16) Pons, S., Bewick, A. Langmuir 1985,
- (17) Bewick, A.; Gibilaro C., Pons, S., submitted for publication in Langmuir.



John Foley received a BS from the University of London in 1981 and an MS from the University of Southampton in 1982. He is currently completing work for a PhD under the direction of Stanley Pons. His research is mainly in the field of IR spectroelectrochemistry.



Stanley Pons received a BS degree from Wake Forest University in 1965 and a PhD from the University of Southampton in 1979. He is currently associate professor of chemistry at the University of Utah, where his research interests include the areas of spectroelectrochemistry, ultramicroelectrodes, electrochemical reaction mechanisms and catalysis, and electrochemical energy sources.

FEMALOS PERSONAL PERSONAL PERSONAL PROPERTY PRODUCES PROPERTY

TECHNICAL REPORT DISTRIBUTION LIST, GEN

<u>c</u>	No. opies		No. <u>Copies</u>
Office of Naval Research Attn: Code 413 800 N. Quincy Street Arlington, Virginia 22217	2	Dr. David Young Code 334 NORDA NSTL, Mississippi 39529	1
Dr. Bernard Douda Naval Weapons Support Center Code 5042 Crane, Indiana 47522	1	Naval Weapons Center Attn: Dr. Ron Atkins Chemistry Division China Lake, California 93555	1
Commander, Naval Air Systems Command Attn: Code 310C (H. Rosenwasser) Washington, D.C. 20360	1	Scientific Advisor Commandant of the Marine Corps Code RD-1 Washington, D.C. 20380	:
Naval Civil Engineering Laboratory Attn: Dr. R. W. Drisko Port Hueneme, California 93401	1	U.S. Army Research Office Attn: CRD-AA-IP P.O. Box 12211 Research Triangle Park, NC 2775	: ;9
Defense Technical Information Center Building 5, Cameron Station Alexandria, Virginia 22314	12	Mr. John Boyle Materials Branch Naval Ship Engineering Center Philadelphia, Pennsylvania 1912	:
DTNSRDC Attn: Dr. G. Bosmajian Applied Chemistry Division Annapolis, Maryland 21401	1	Naval Ocean Systems Center Attn: Dr. S. Yamamoto Marine Sciences Division San Diego, California 91232	1
Dr. William Tolles Superintendent Chemistry Division, Code 6100 Naval Research Laboratory Washington, D.C. 20375	1		

Dr. Paul Delahay Department of Chemistry New York University New York, New York 10003

Dr. P. J. Hendra
Department of Chemistry
University of Southampton
Southampton S09 5NH
United Kingdom

Dr. J. Driscoll Lockheed Palo Alto Research Laboratory 3251 Hanover Street Palo Alto, California 94304

Dr. D. N. Bennion
Department of Chemical Engineering
Brigham Young University
Provo, Utah 84602

Dr. R. A. Marcus Department of Chemistry California Institute of Technology Pasadena, California 91125

Dr. J. J. Auborn Bell Laboratories Murray Hill, New Jersey 07974

Dr. Joseph Singer, Code 302-1 NASA-Lewis 21000 Brookpark Road Cleveland, Ohio 44135

Dr. P. P. Schmidt
Department of Chemistry
Oakland University
Rochester, Michigan 48063

Dr. Manfred Breiter
Institut fur Technische Elektrochemie
Technischen Universitat Wien
9 Getreidemarkt, 1160 Wien
AUSTRIA

Dr. E. Yeager Department of Chemistry Case Western Reserve University Cleveland, Ohio 44106

Dr. C. E. Mueller
The Electrochemistry Branch
Naval Surface Weapons Center
White Oak Laboratory
Silver Spring, Maryland 20910

Dr. Sam Perone
Chemistry & Materials
Science Department
Lawrence Livermore National Laboratory
Livermore, California 94550

Dr. Royce W. Murray
Department of Chemistry
University of North Carolina
Chapel Hill, North Carolina 27514

Dr. B. Brummer EIC Incorporated 111 Downey Street Norwood, Massachusetts 02062

Dr. Adam Heller Bell Laboratories Murray Hill, New Jersey 07974

Dr. A. B. Ellis Chemistry Department University of Wisconsin. Madison, Wisconsin 53706

የ**ፅመጀመን የመጀመሪ መ**ጀመሪ እንደ ተርፈጥር እና ለተርፈጥር እርስ መመር ለተርፈጥር እና ለተርፈጥር እና ለተርፈጥር እና ለተርፈጥር እና ለተርፈጥር እና ለተርፈጥር እና ለተርፈጥር

Library
Duracell, Inc.
Burlington, Massachusetts 01803

Electrochimica Corporation 20 Kelly Court Menlo Park, California 94025-1418

Dr. M. Wrighton
Chemistry Department
Massachusetts Institute
of Technology
Cambridge, Massachusetts 02139

Dr. B. Stanley Pons
Department of Chemistry
University of Utah
Salt Lake City, Utah 84112

Donald E. Mains
Naval Weapons Support Center
Electrochemical Power Sources Division
Crane, Indiana 47522

S. Ruby
DOE (STOR)
Room 5E036 Forrestal Bldg., CE-14
Washington, D.C. 20595

Dr. A. J. Bard
Department of Chemistry
University of Texas
Austin, Texas 78712

PARKETON TOUCHOUSE IN DOCUMENT OF THE PARKETON TO THE PARKET OF THE PARK

Or. Janet Osteryoung Department of Chemistry State University of New York Buffalo, New York 14214

Dr. Donald W. Ernst Naval Surface Weapons Center Code R-33 White Oak Laboratory Silver Spring, Maryland 20910

Mr. James R. Moden Naval Underwater Systems Center Code 3632 Newport, Rhode Island 02840

Dr. Bernard Spielvogel U.S. Army Research Office P.O. Box 12211 Research Triangle Park, NC 27709

Or. Aaron Fletcher Naval Weapons Center Code 3852 China Lake, California 93555 Dr. M. M. Nicholson Electronics Research Center Rockwell International 3370 Miraloma Avenue Anaheim, California

Dr. Michael J. Weaver
Department of Chemistry
Purdue University
West Lafayette, Indiana 47907

Dr. R. David Rauh EIC Laboratories, Inc. 111 Downey Street Norwood, Massachusetts 02062

Dr. Aaron Wold Department of Chemistry Brown University Providence, Rhode Island 02192

Dr. Martin Fleischmann Department of Chemistry University of Southampton Southampton SO9 5NH ENGLAND

Dr. R. A. Osteryoung Department of Chemistry State University of New York Buffalo, New York 14214

Dr. John Wilkes Air Force Office of Scientific Research Bolling AFB Washington, D.C. 20332

Dr. R. Nowak Naval Research Laboratory Code 6171 Washington, D.C. 20375

Dr. D. F. Shriver
Department of Chemistry
Northwestern University
Evanston, Illinois 60201

Dr. Hector D. Abruna Department of Chemistry Cornell University Ithaca. New York 14853

Dr. A. B. P. Lever Chemistry Department York University Downsview, Ontario M3J1P3

Dr. Stanislaw Szpak Naval Ocean Systems Center Code 633, Bayside San Diego, California 95152

Dr. Gregory Farrington
Department of Materials Science
and Engineering
University of Pennsylvania
Philadelphia, Pennsylvania 19104

M. L. Robertson
Manager, Electrochemical
and Power Sources Division
Naval Weapons Support Center
Crane, Indiana 47522

Dr. T. Marks
Department of Chemistry
Northwestern University
Evanston, Illinois 60201

Dr. Micha Tomkiewicz Department of Physics Brooklyn College Brooklyn, New York 11210

Dr. Lesser Blum
Department of Physics
University of Puerto Rico
Rio Piedras, Puerto Rico 00931

Dr. Joseph Gordon, II IBM Corporation 5600 Cottle Road San Jose. California 95193

Dr. Nathan Lewis Department of Chemistry Stanford University Stanford, California 94305 Dr. D. H. Whitmore
Department of Materials Science
Northwestern University
Evanston, Illinois 60201

Dr. Alan Bewick Department of Chemistry The University of Southampton Southampton, SO9 5NH ENGLAND

Dr. E. Anderson NAVSEA-56Z33 NC #4 2541 Jefferson Davis Highway Arlington, Virginia 20362

Dr. Bruce Dunn
Department of Engineering &
Applied Science
University of California
Los Angeles, California 90024

Dr. Elton Cairns
Energy & Environment Division
Lawrence Berkeley Laboratory
University of California
Berkeley, California 94720

Dr. Richard Pollard Department of Chemical Engineering University of Houston Houston, Texas 77004

Dr. M. Philpott IBM Corporation 5600 Cottle Road San Jose, California 95193

Dr. Donald Sandstrom Boeing Aerospace Co. P.O. Box 3999 Seattle, Washington 98124

Dr. Carl Kannewurf
Department of Electrical Engineering
and Computer Science
Northwestern University
Evanston, Illinois 60201

Dr. Joel Harris
Department of Chemistry
University of Utah
Salt Lake City, Utah 84112

ቜ**ቘቔቔ፠**ዄቔፚቔጜዄቒዸቒዸቒ፟ቒቒቜዸኇጜጜቔዸዸጜኯ፟፟፟፟ፘኯፙኯጜጜዀዀቜቜቜቜቔዀዀኯፙኯፙኯፙኯፙጜፙኯፙቔፙኯፙኯፙኯፙኯፙኯፙኯዀኯኯኯኯቜቔጜቜቜቜ

Dr. Robert Somoano Jet Propulsion Laboratory California Institute of Technology Pasadena, California 91103

Dr. Johann A. Joebstl
USA Mobility Equipment R&D Command
DRDME-EC
Fort Belvoir, Virginia 22060

Dr. Judith H. Ambrus NASA Headquarters M.S. RTS-6 Washington, D.C. 20546

Dr. Albert R. Landgrebe U.S. Department of Energy M.S. 68025 Forrestal Building Washington, D.C. 20595

Dr. J. J. Brophy
Department of Physics
University of Utah
Salt Lake City, Utah 84112

Dr. Charles Martin
Department of Chemistry
Texas A&M University
College Station, Texas 77843

Dr. H. Tachikawa Department of Chemistry Jackson State University Jackson, Mississippi 39217

Dr. Theodore Beck Electrochemical Technology Corp. 3935 Leary Way N.W. Seattle, Washington 98107

Dr. Farrell Lytle
Boeing Engineering and
Construction Engineers
P.O. Box 3707
Seattle, Washington 98124

Dr. Robert Gotscholl U.S. Department of Energy MS G-226 Washington, D.C. 20545 Dr. Edward Fletcher Department of Mechanical Engineering University of Minnesota Minneapolis, Minnesota 55455

Dr. John Fontanella Department of Physics U.S. Naval Academy Annapolis, Maryland 21402

Dr. Martha Greenblatt
Department of Chemistry
Rutgers University
New Brunswick, New Jersey 08903

Dr. John Wasson Syntheco, Inc. Rte 6 - Industrial Pike Road Gastonia. North Carolina 28052

Dr. Walter Roth
Department of Physics
State University of New York
Albany, New York 12222

Dr. Anthony Sammells Eltron Research Inc. 4260 Westbrook Drive, Suite 111 Aurora, Illinois 60505

Dr. C. A. Angell Department of Chemistry Purdue University West Lafayette, Indiana 47907

Dr. Thomas Davis
Polymer Science and Standards
Division
National Bureau of Standards
Washington, D.C. 20234

Ms. Wendy Parkhurst
Naval Surface Weapons Center R-33
R-33
Silver Spring, Maryland 20910

Dr. John Owen
Department of Chemistry and
Applied Chemistry
University of Salford
Salford M5 4WT ENGLAND

Dr. Boone Owens
Department of Chemical Engineering
and Materials Science
University of Minnesota
Minneapolis, Minnesota 55455

Dr. J. O. Thomas University of Uppsala Institute of Chemistry Box 531 S-751 21 Uppsala, Sweden Dr. O. Stafsudd Department of Electrical Engineering University of California Los Angeles, California 90024

Dr. S. G. Greenbaum Department of Physics Hunter College of CUNY New York, New York 10021

Dr. Menahem Anderman W.R. Grace & Co. Columbia, Maryland 20144

END 1) A TE FILMED 6-1988 DTIC